

# Dai meccanismi molecolari allo sviluppo di strategie terapeutiche innovative per i difetti genetici della coagulazione

Francesco Bernardi  
Università di Ferrara



**SISET  
2016**

## Molecular mechanisms for new therapeutic approaches (1)

### Substitutive therapy (protein)

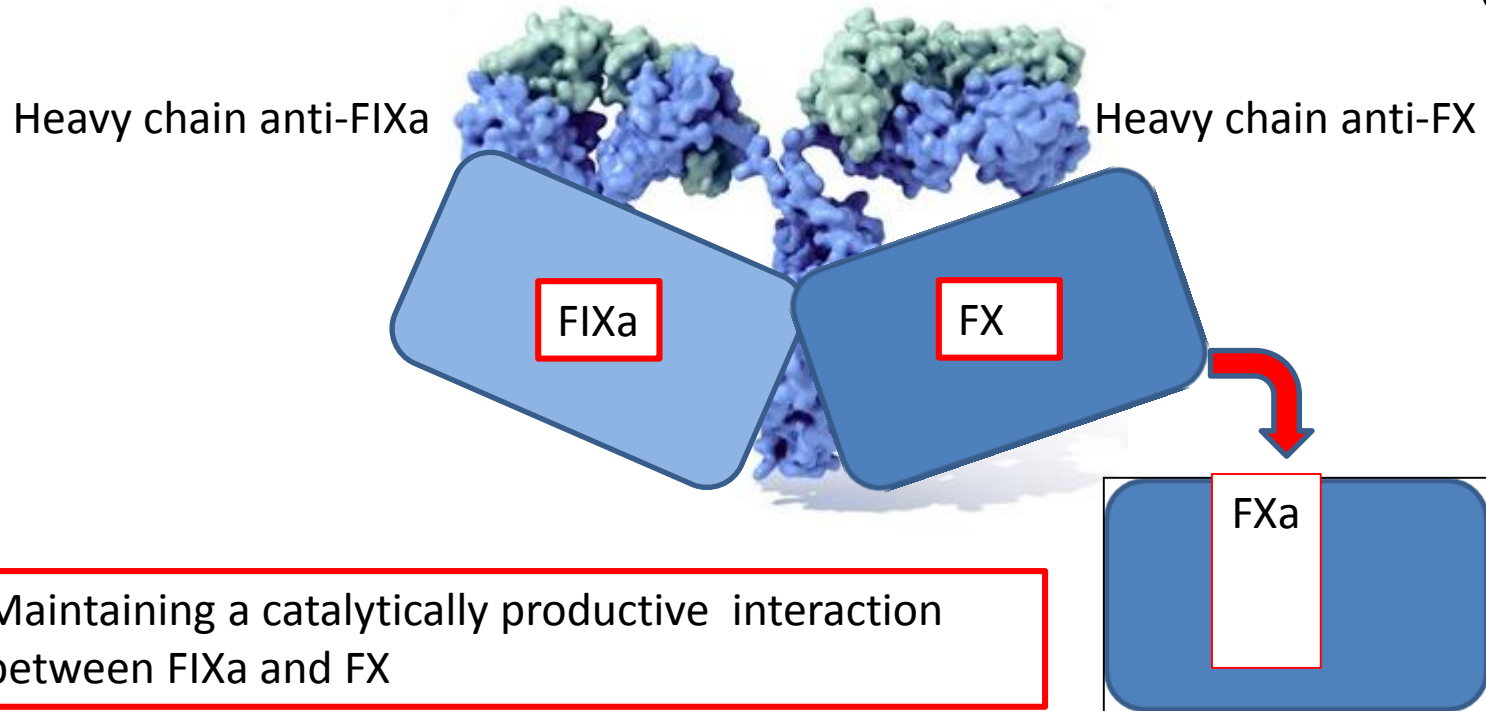
- extending the half-life/bio-distribution of circulating exogenous factor protein
- mimicking the coagulation function of factor VIII by bispecific antibody technology

### Disrupting anticoagulant proteins (antibodies, aptamers, RNAi)

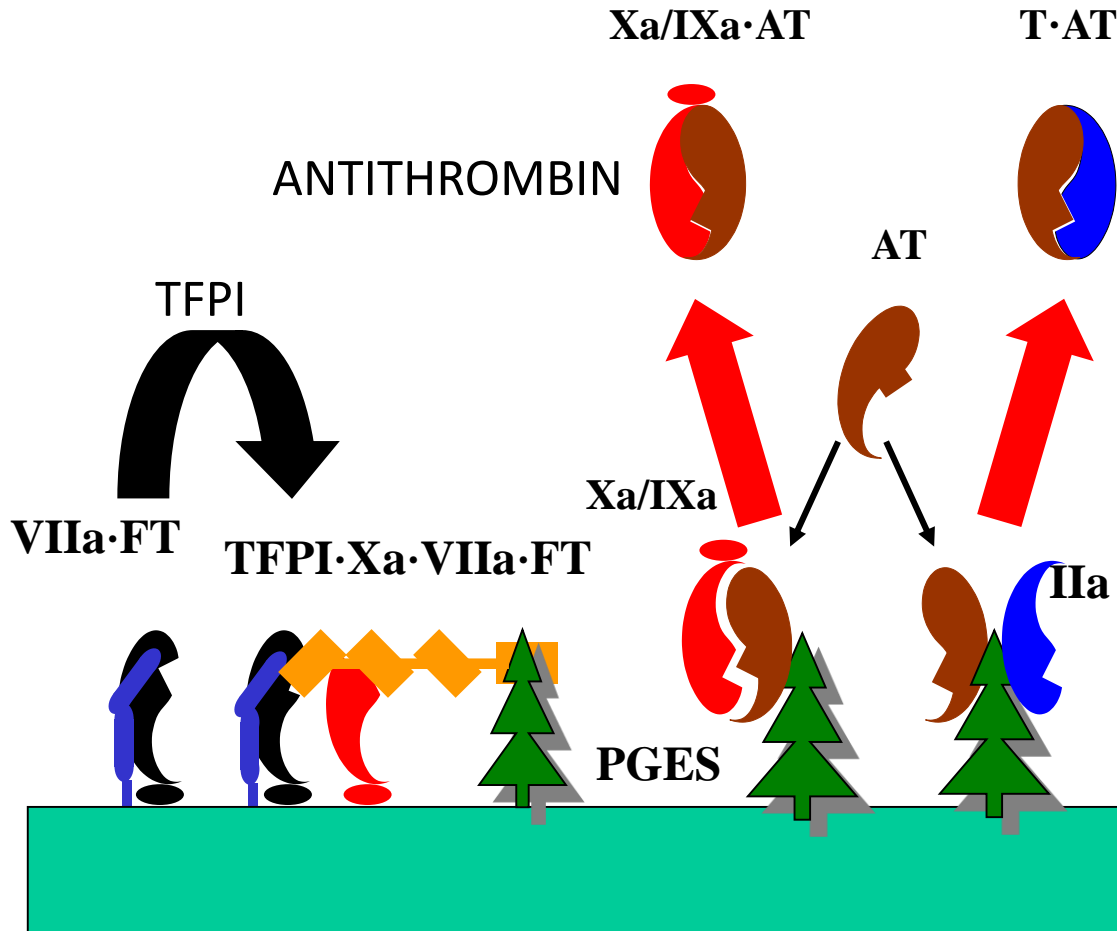
- tissue factor pathway inhibitor (TFPI)
- antithrombin (AT3)

# Humanized asymmetric antibody mimicking FVIIIa function

Emicizumab (ACE910)

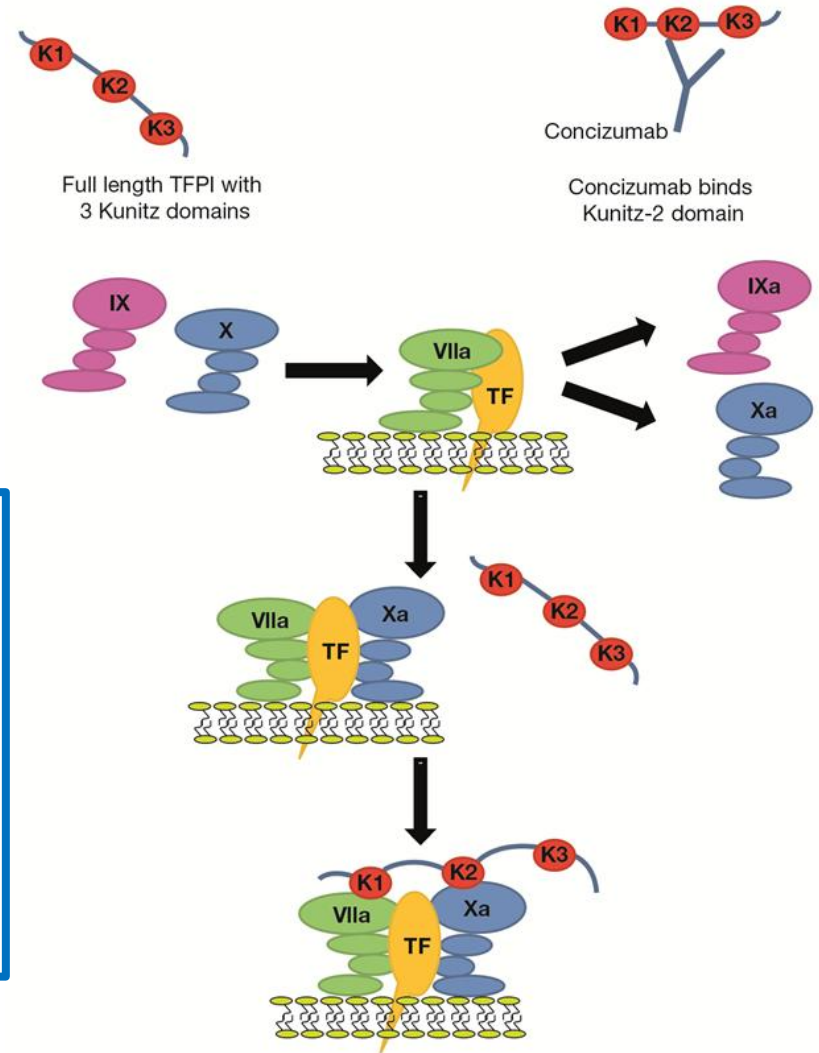
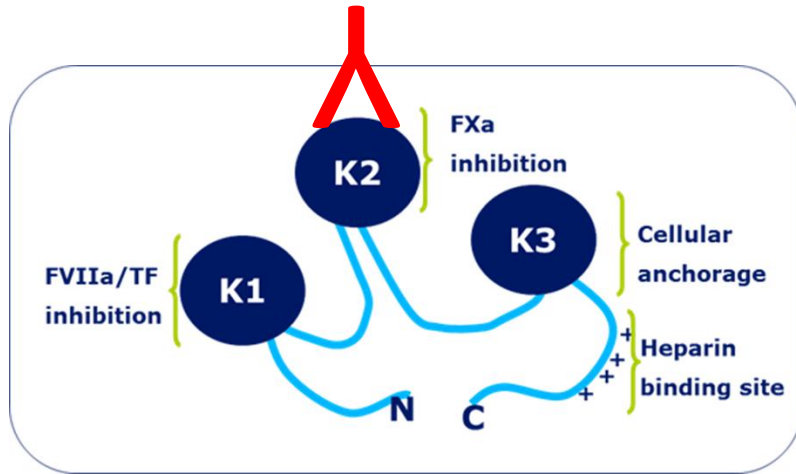


# Decreasing anticoagulant function: Inhibition of inhibitors



TOOLS  
Antibodies  
Aptamers  
RNAi

# Inhibition of TFPI by specific antibodies (Concizumab)

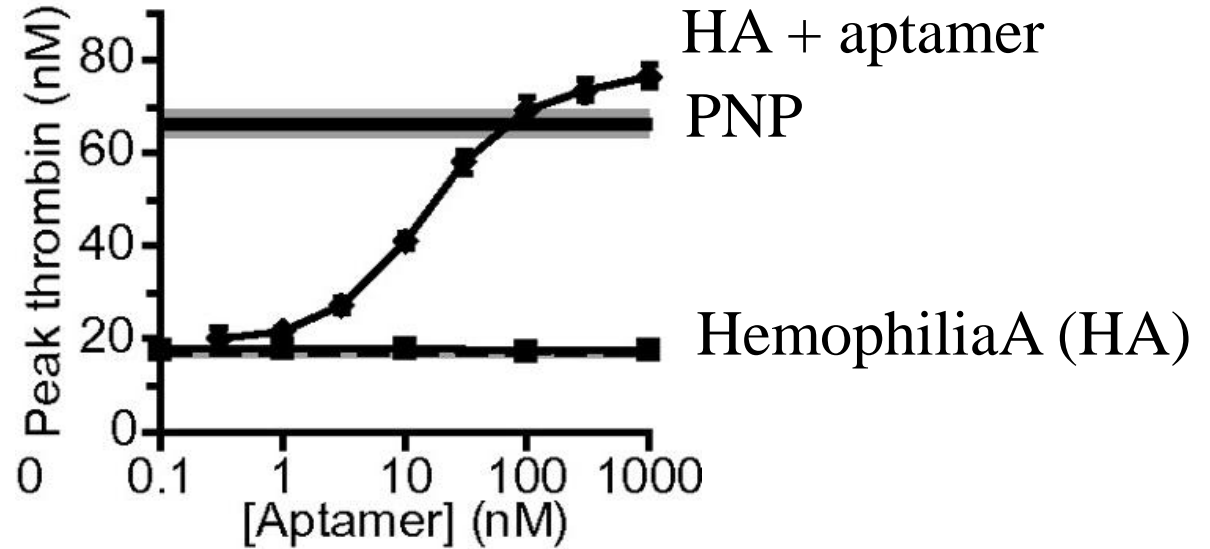


Blockage of the KPI-2 domain prevents TFPI binding to both FXa and FVIIa/TF

Downregulation of TFPI inhibition of the coagulation cascade

Further FXa generation in response to tissue injury.

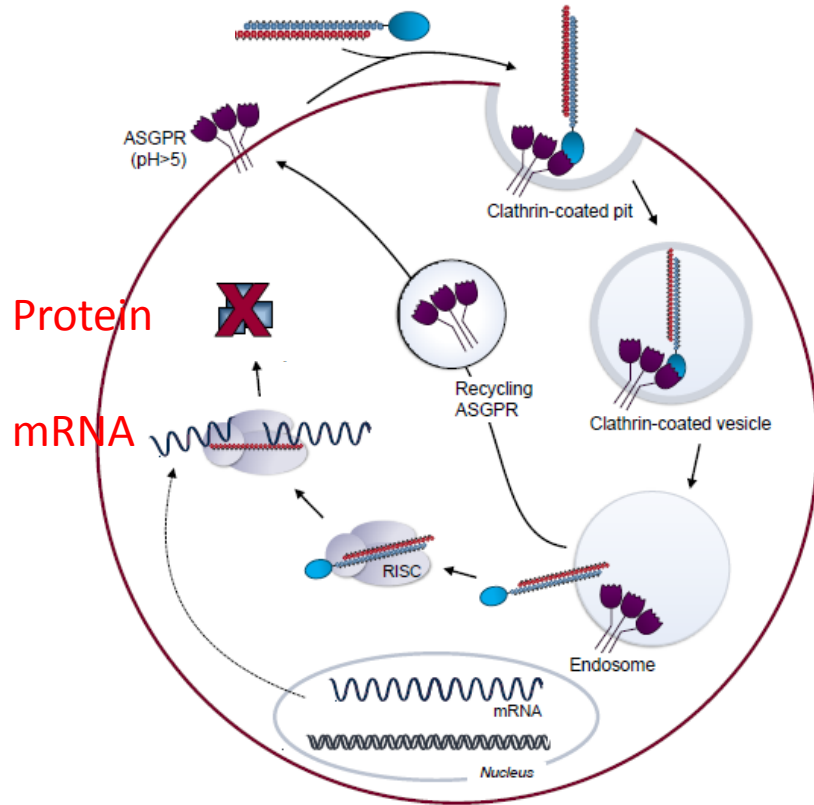
**Aptamer anti TFPI ARC19499:  
increased thrombin generation in Hemophilia plasma**



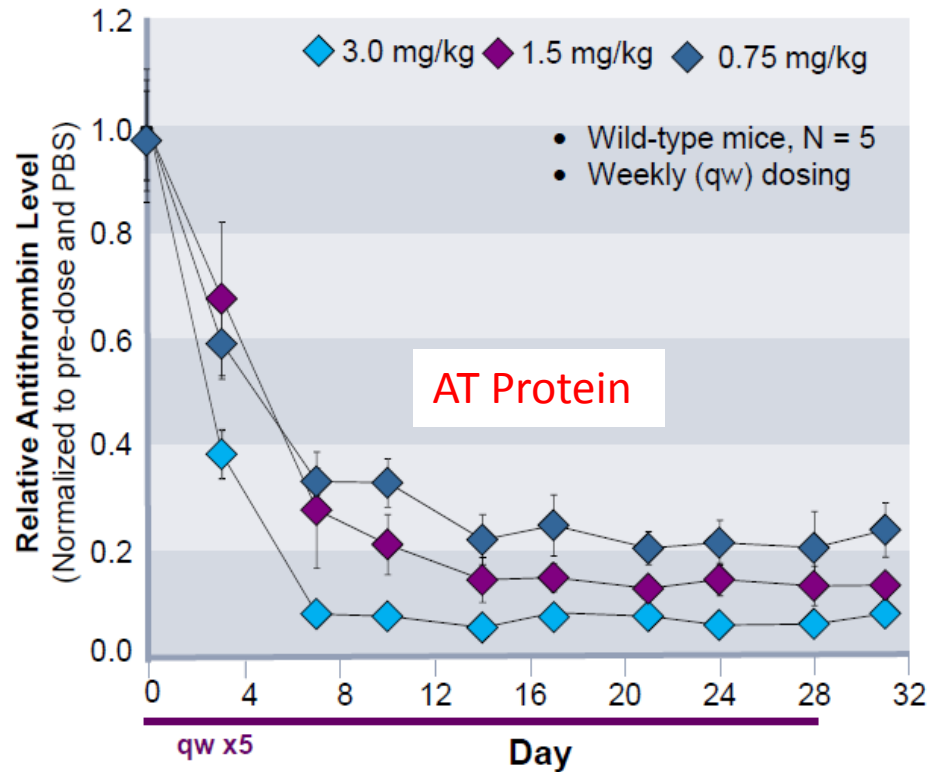
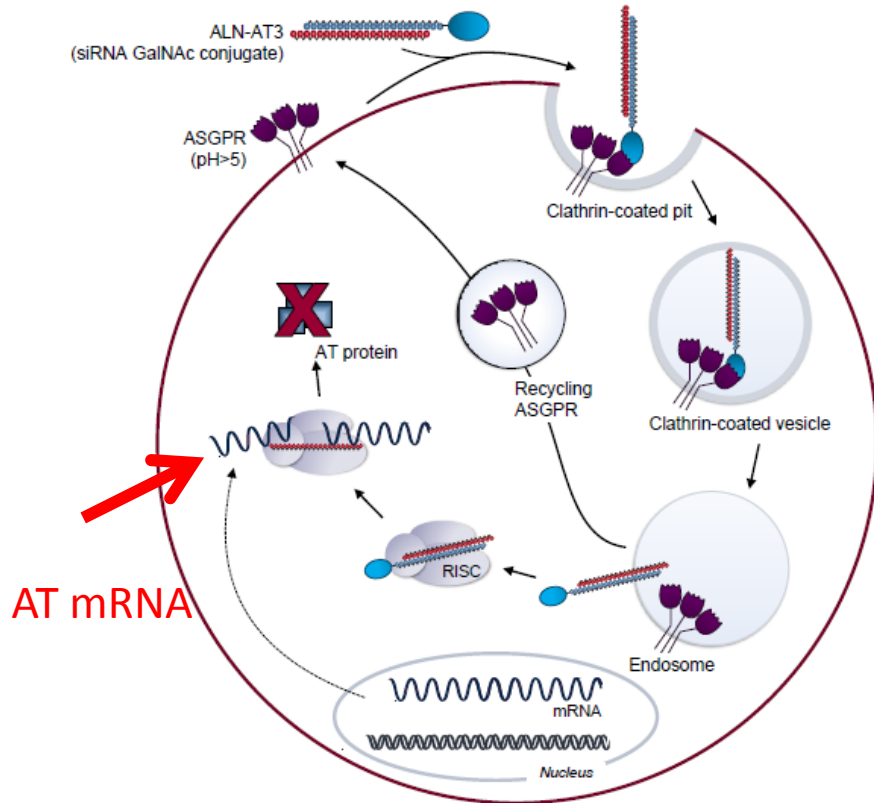
ARC19499 (◆)  
negative control oligonucleotide (■).

Waters E K et al. Blood 2011;117:5514-5522

# RNA interference (RNAi)



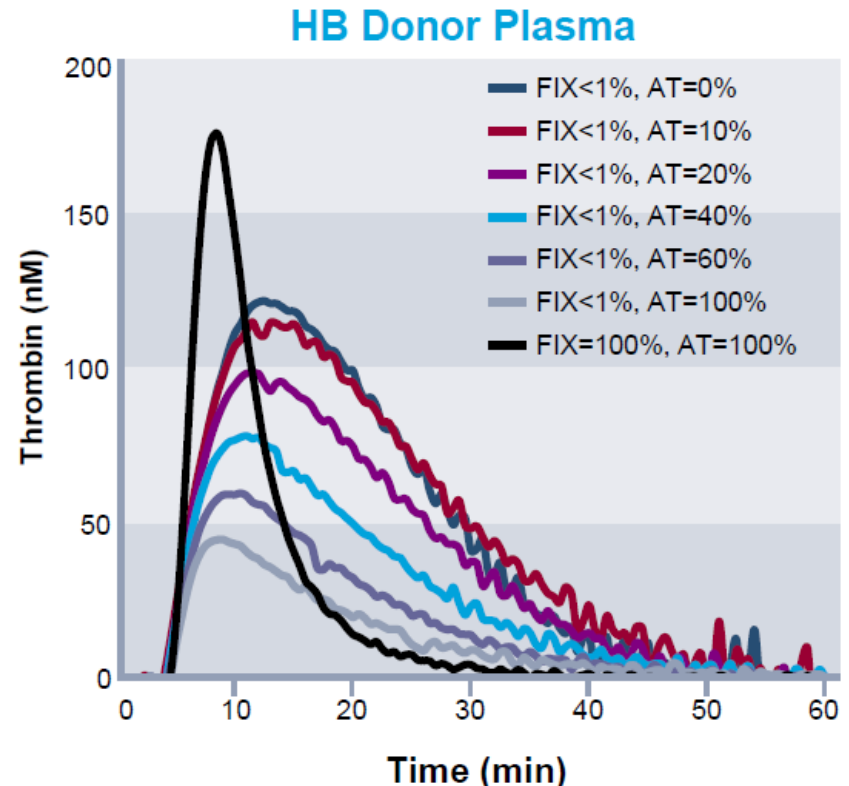
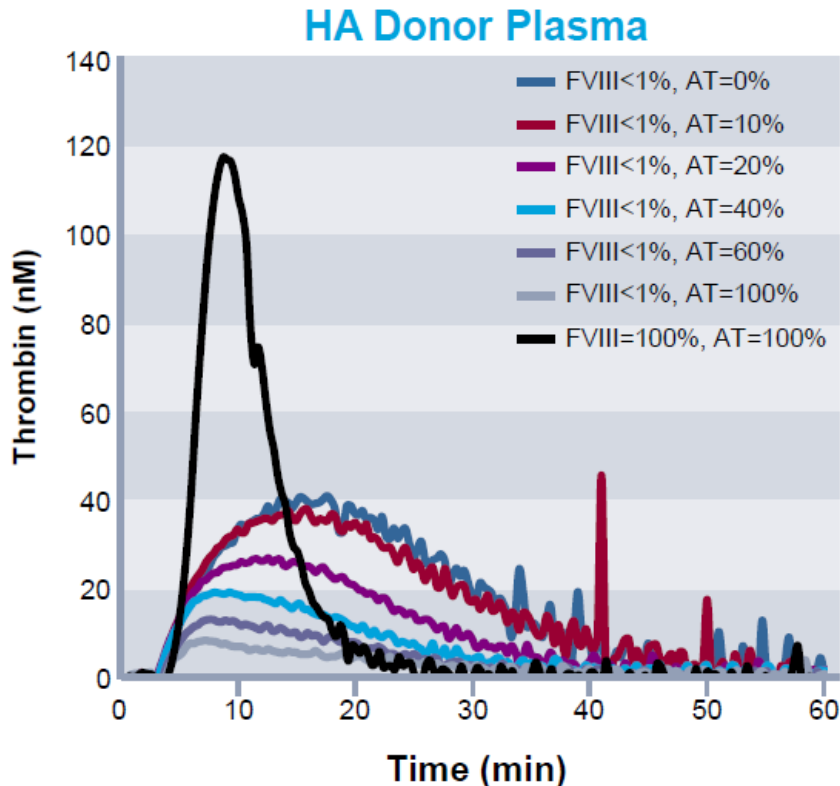
# RNAi Targeting Antithrombin





# Antithrombin Depletion Increased Thrombin Generation in Hemophilia Plasma

ALN-AT3



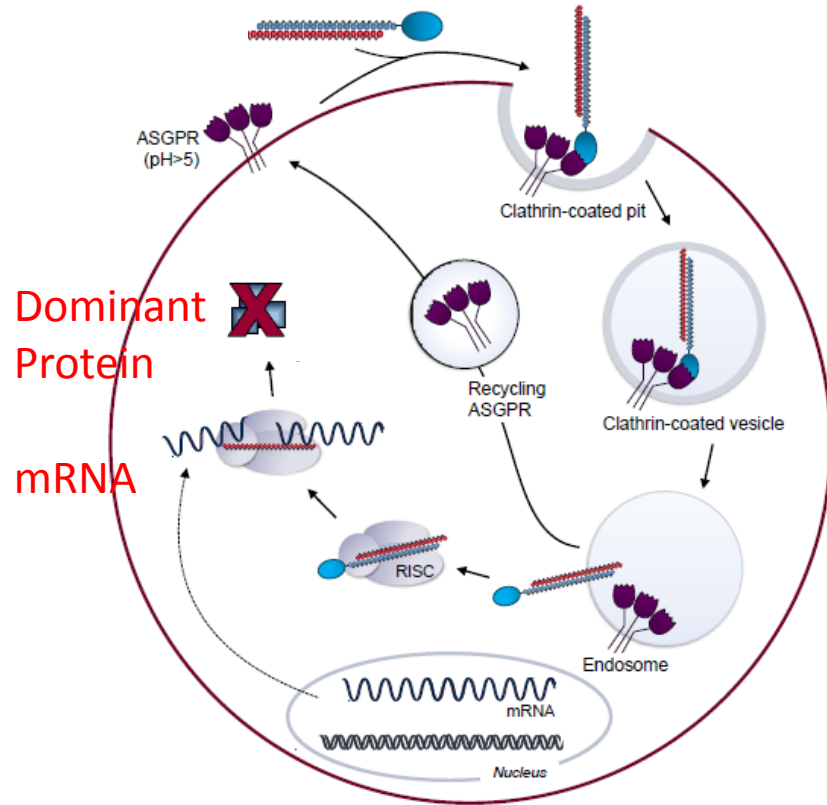
- Antithrombin depletion increases peak height and delays inhibition of thrombin

## Molecular mechanisms for new therapeutic approaches (2)

### RNA directed/based therapy

- Engineered small RNA: RNAi, U1snRNA
- Drug-induced translational readthrough of stop codons
- Transcription activation TALE-TF

# Counteracting the dominant negative effect in VWD type 2 by RNAi



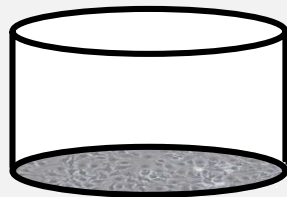
Caterina Casari

# Counteracting the dominant-negative effect

*in vitro*

Plasmids  
pSV-hVWF-DEL  
pSV-hVWF-wt

*Co-transfection*



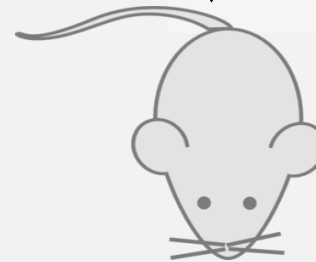
COS-1 cells

Heterozygous state  
expression

*in vivo*

Plasmids  
pLIVE-mVWF-DEL  
pLIVE-mVWF-wt

*Co-injection via  
hydrodynamic  
gene transfer*



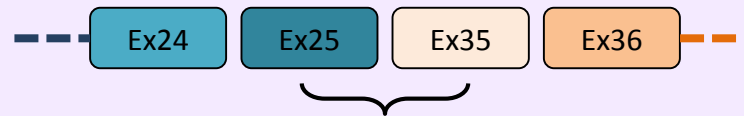
Vwf<sup>-/-</sup> mice

VWF antigen levels & VWF multimer profile were assessed in conditioned media and in plasma.

Bleeding phenotype was assessed in vivo

# Counteracting the dominant-negative effect: allele-specific RNAi *-in vitro-*

- Use RNAi to specifically block expression of the dominant negative allele



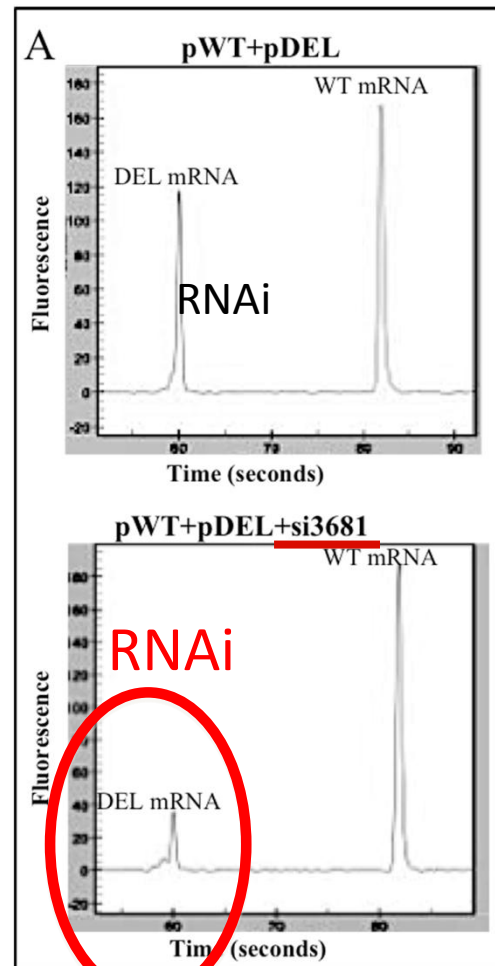
Mutant junction sequence  
connecting exons 25-35 is  
targeted by RNAi

WT-VWF not affected

# Counteracting the dominant-negative effect: allele-specific RNAi *-in vitro-*

- Use RNAi to specifically block expression of the dominant negative allele

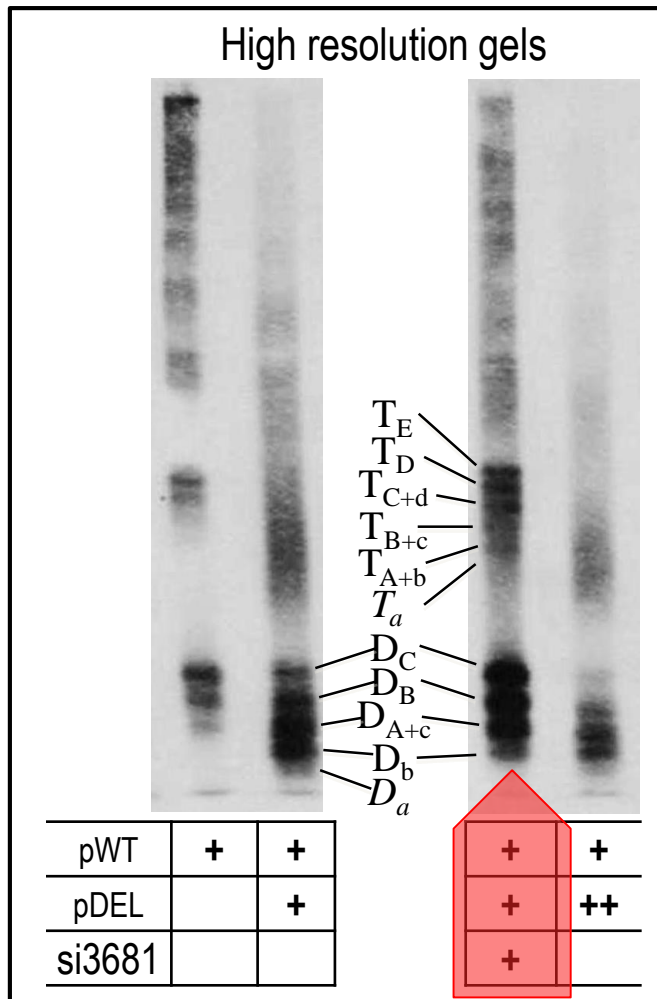
si3681 decreased  
DEL-VWF mRNA  
&  
did not affect  
WT-VWF mRNA



Capillary electrophoresis  
of RT-PCR products  
from transfected cells

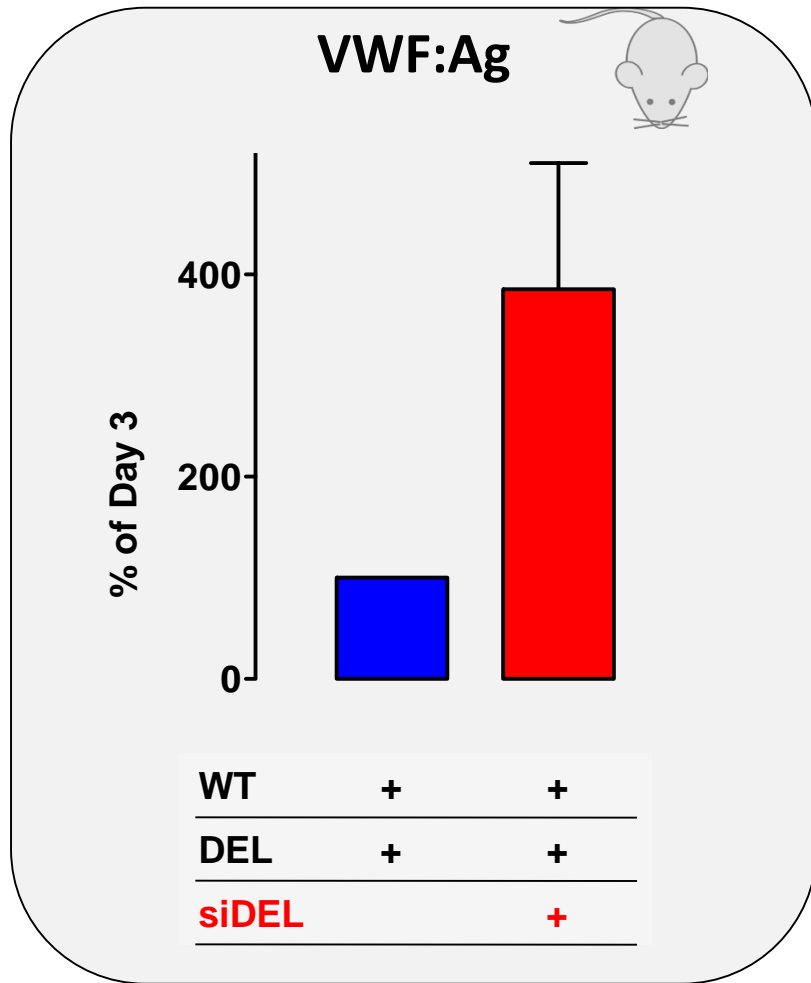
Selective decrease  
of the altered mRNA  
and thus of the  
dominant negative  
VWF

# Counteracting the dominant-negative effect: Partial rescue of VWF



Partial rescue  
of HMWMs

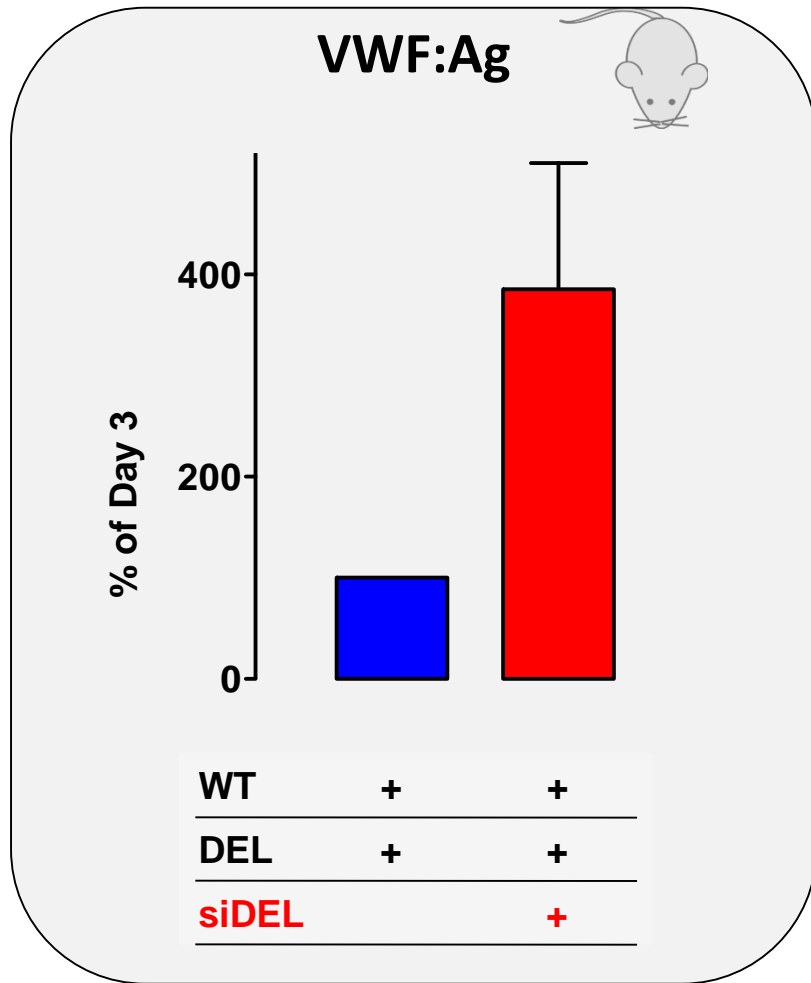
# *in vivo* RNA interference



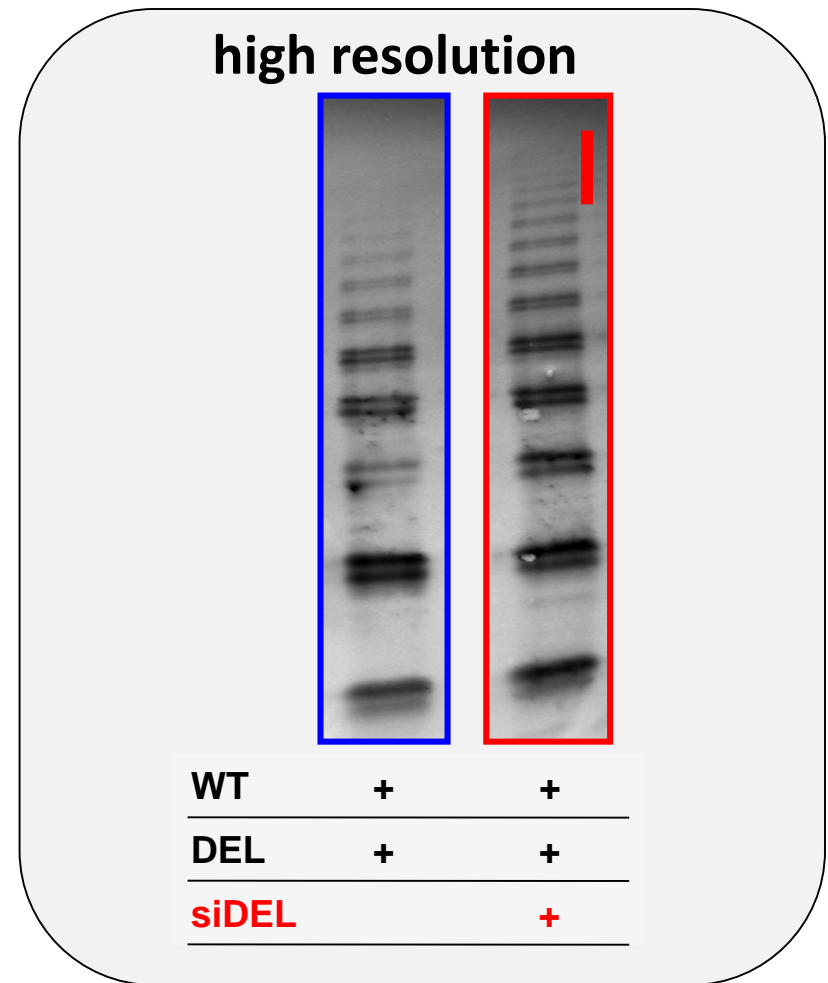
3.8 fold increase of the antigen levels by siDEL administration



# *in vivo* RNA interference



3.8 fold increase of the antigen levels by siDEL administration



improvement of the multimer profile

# Counteracting the dominant-negative effect in VWD type 2 by RNAi

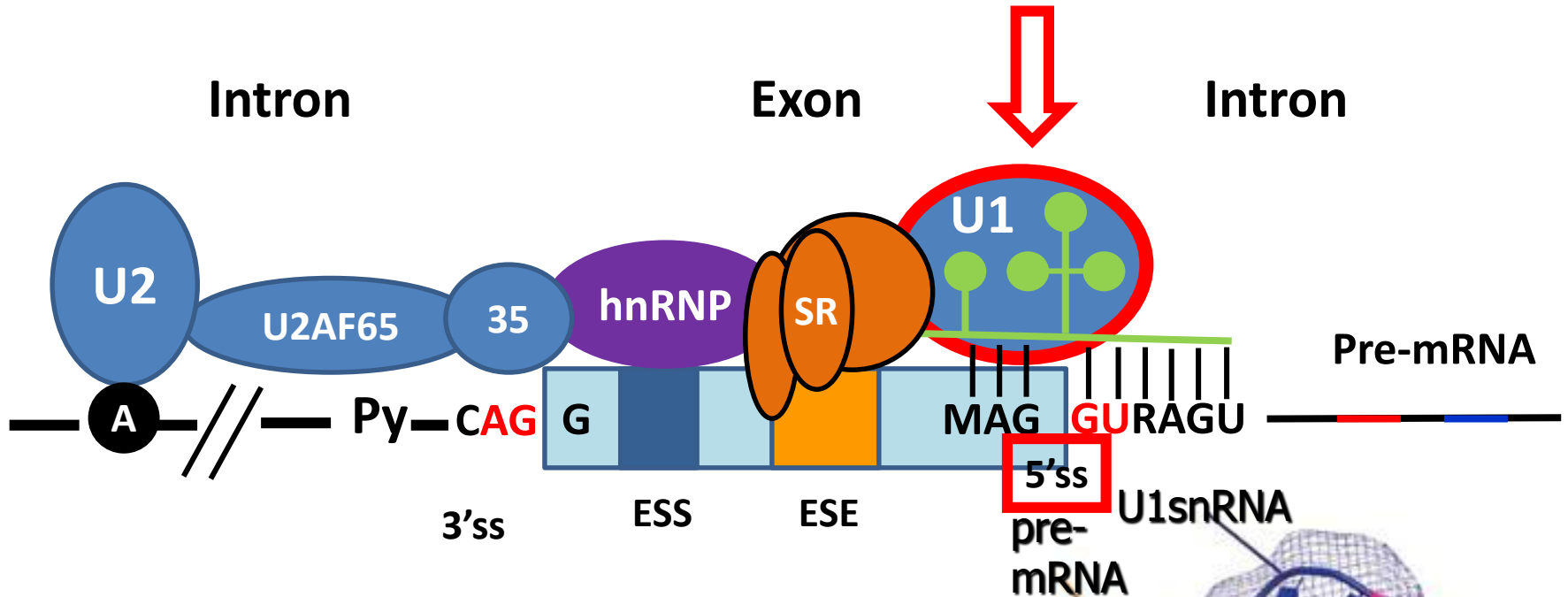
- The effects of mutations with dominant-negative effect were reproduced for the first time *in vivo* in *Vwf*<sup>-/-</sup> mice
- In our mouse model for the heterozygous expression of mutations with dominant-negative effects on VWF, we were able to counteract the deleterious effects of the mutations by the *in vivo* administration of siRNAs, selectively targeting the mutant mRNA molecules
  - *siRNA was administered for the first time in a mouse model of VWD and could represent a therapeutic approach*

## Molecular mechanisms for new therapeutic approaches (2)

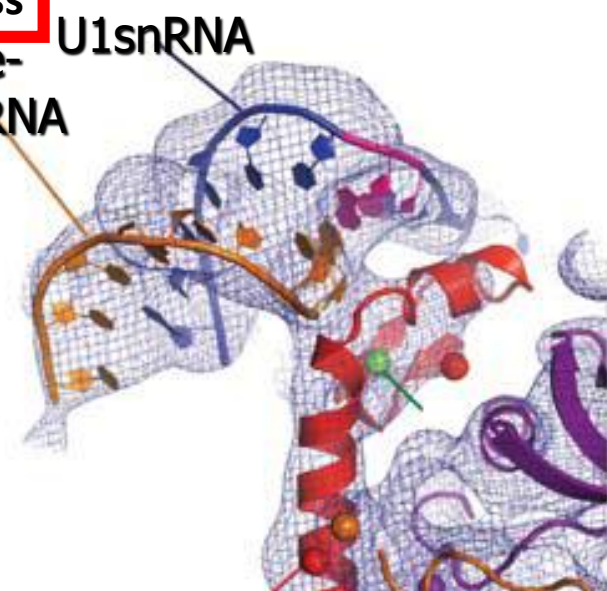
### RNA directed/based therapy

- Engineered small RNA: RNAi, **U1snRNA**
- Drug-induced translational readthrough of stop codons
- Transcription activation TALE-TF

# Splicing steps: focus on U1snRNA



Exon definition

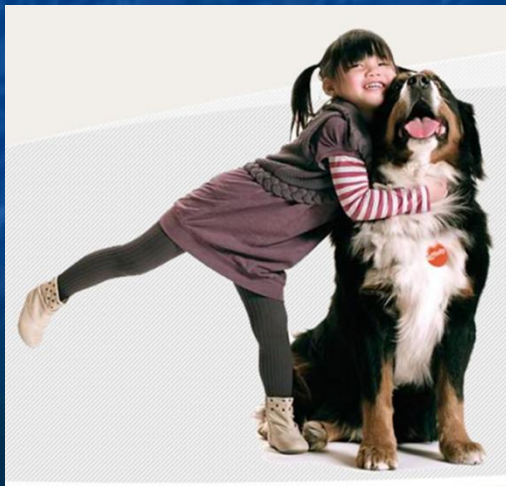


# Recognition of 5'ss by base-pairing to the 5' end of U1 snRNA: Consensus sequence of 5'ss

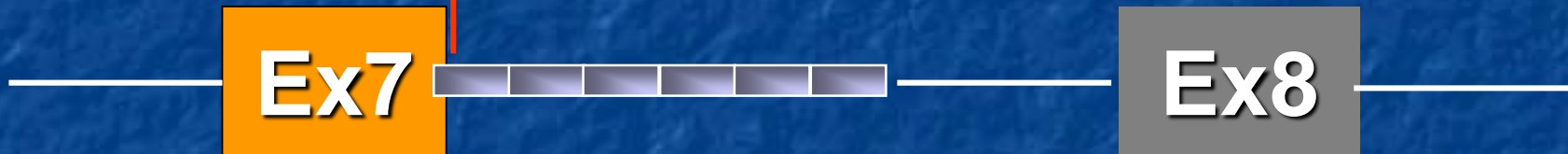
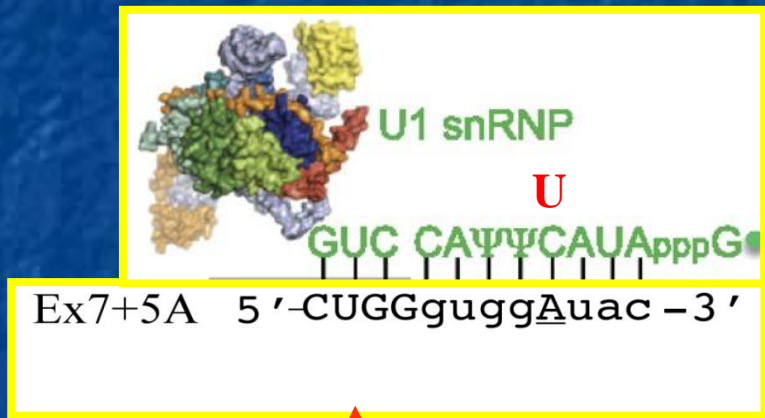
Adapted from Roca X et al. Genes Dev. 2013;27:129-144



## Affinity and position

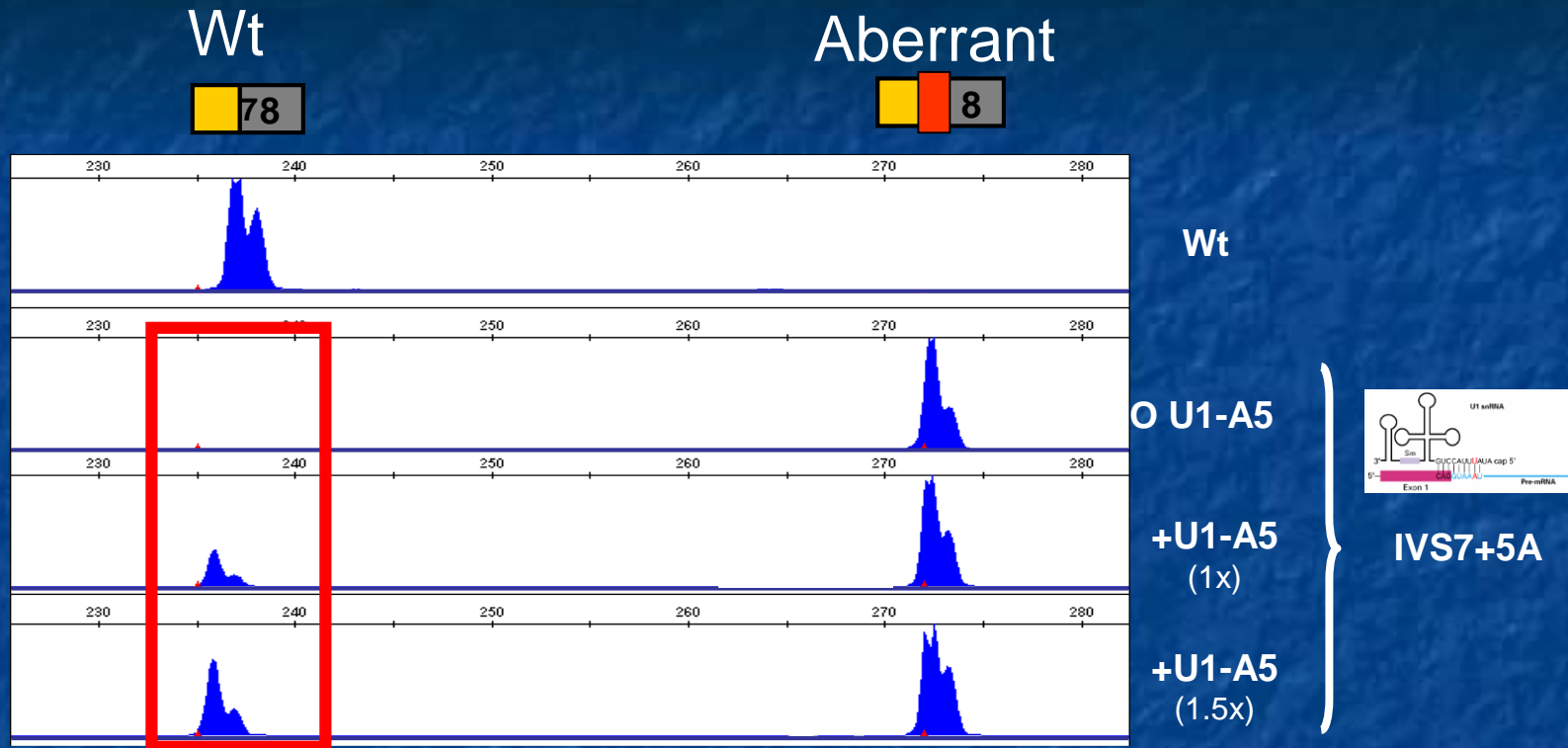


# Splicing Modulation by modified U1 as tools for therapeutic approach



Rescue by Engineered U1snRNA with improved complementarity –increased affinity - to the mutated donor splice site?

# Dose dependent induction of correct splicing

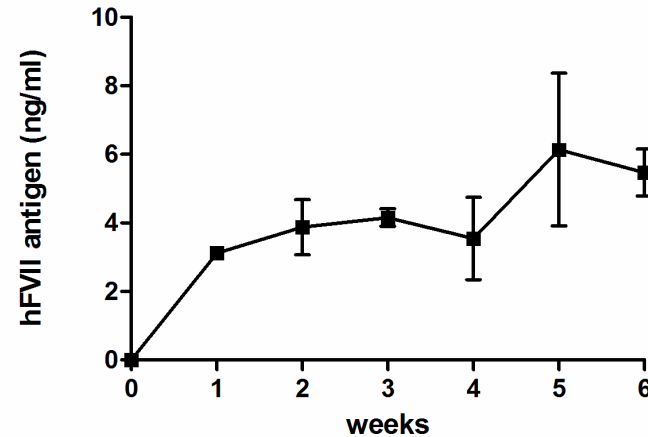


The correctly spliced form is 15±5% of the aberrant form

# The U1+5a-mediated rescue of circulating hFVII levels was prolonged overtime by AAV delivery



Human FVII



|   | AAV2-FVII+5A<br>(vg/mouse) | AAV8-U1+5a<br>(vg/mouse) |
|---|----------------------------|--------------------------|
| ■ | $1.2 \times 10^{12}$       | $1.2 \times 10^{11}$     |





# Splicing Modulation by modified U1

**Mutation** specific U1snRNAs  
Able to correct a single 5'ss change



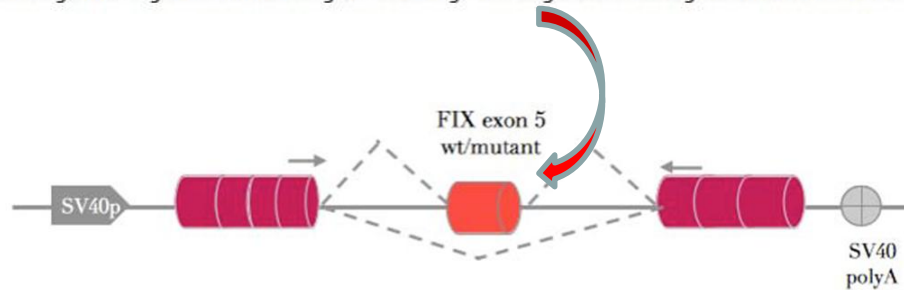
**Exon** specific U1 snRNAs  
able to correct multiple splicing mutations



# Engineered U1s (ExonSpecificU1s) for splicing correction of FIX exon 5 natural mutations

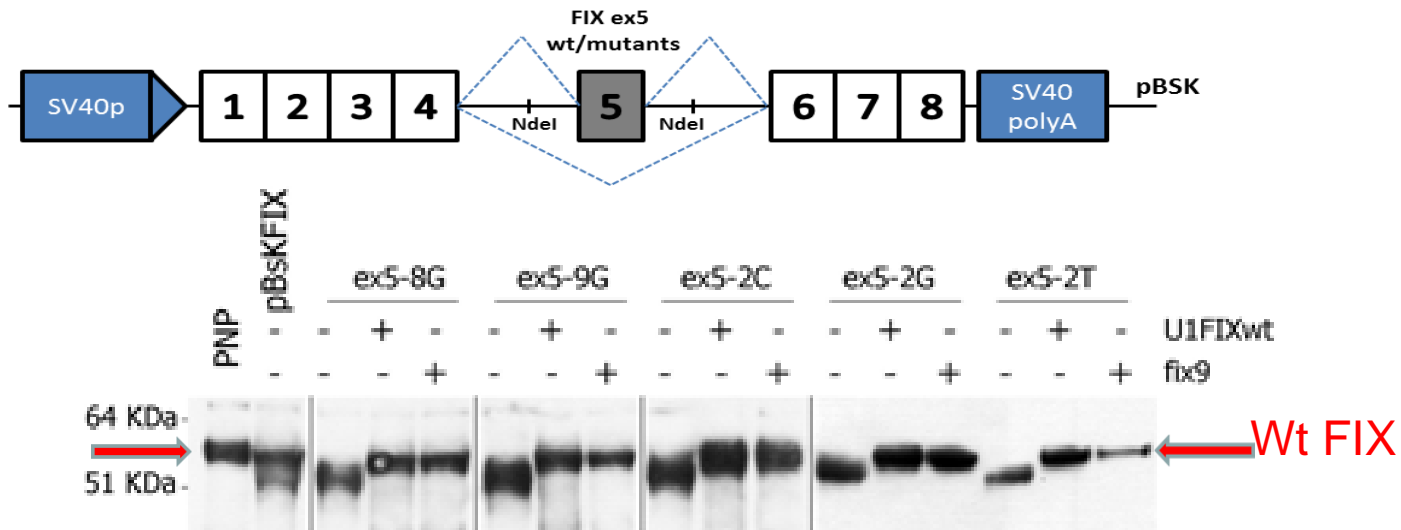


A



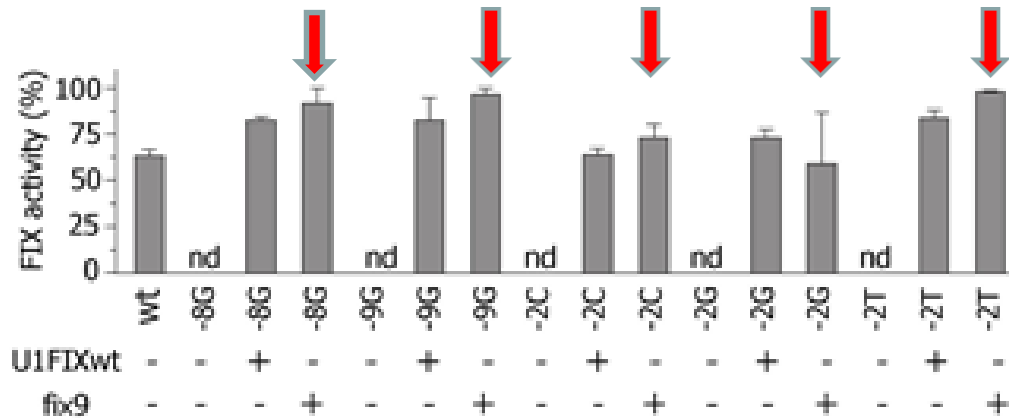
- Targeting U1 snRNAs to specific intronic regions located downstream DDSS to improve exon recognition and inclusion?

# Rescue of FIX protein and FIX function by ExonSpecificU1



Protein

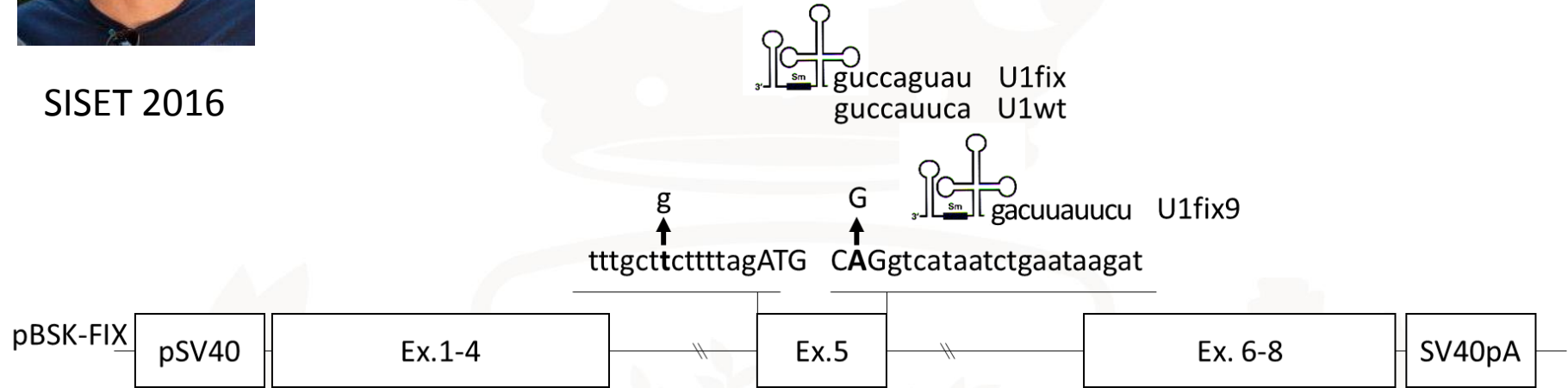
Function



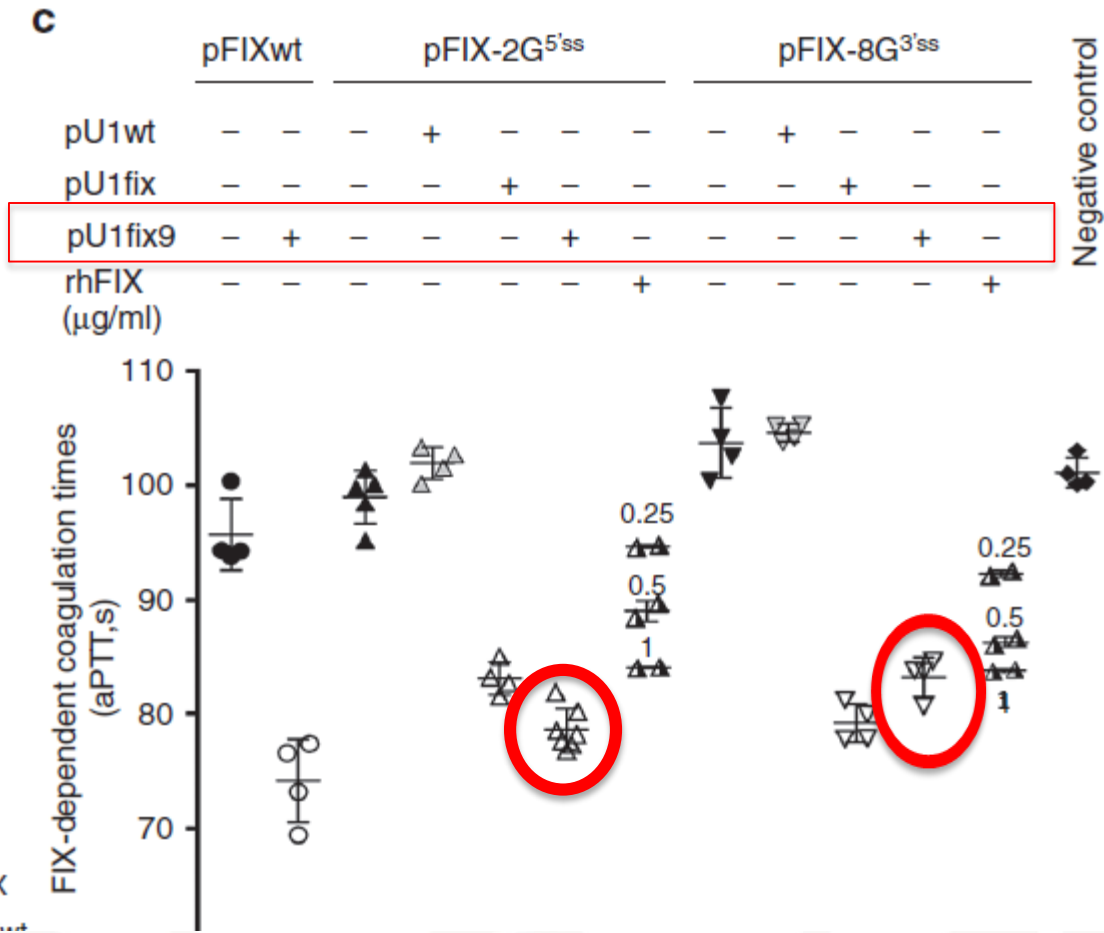


SISSET 2016

# Evaluation of ExSpeU1-mediated rescue



# ExSpeU1-mediated rescue of hFIX expression: coagulation time



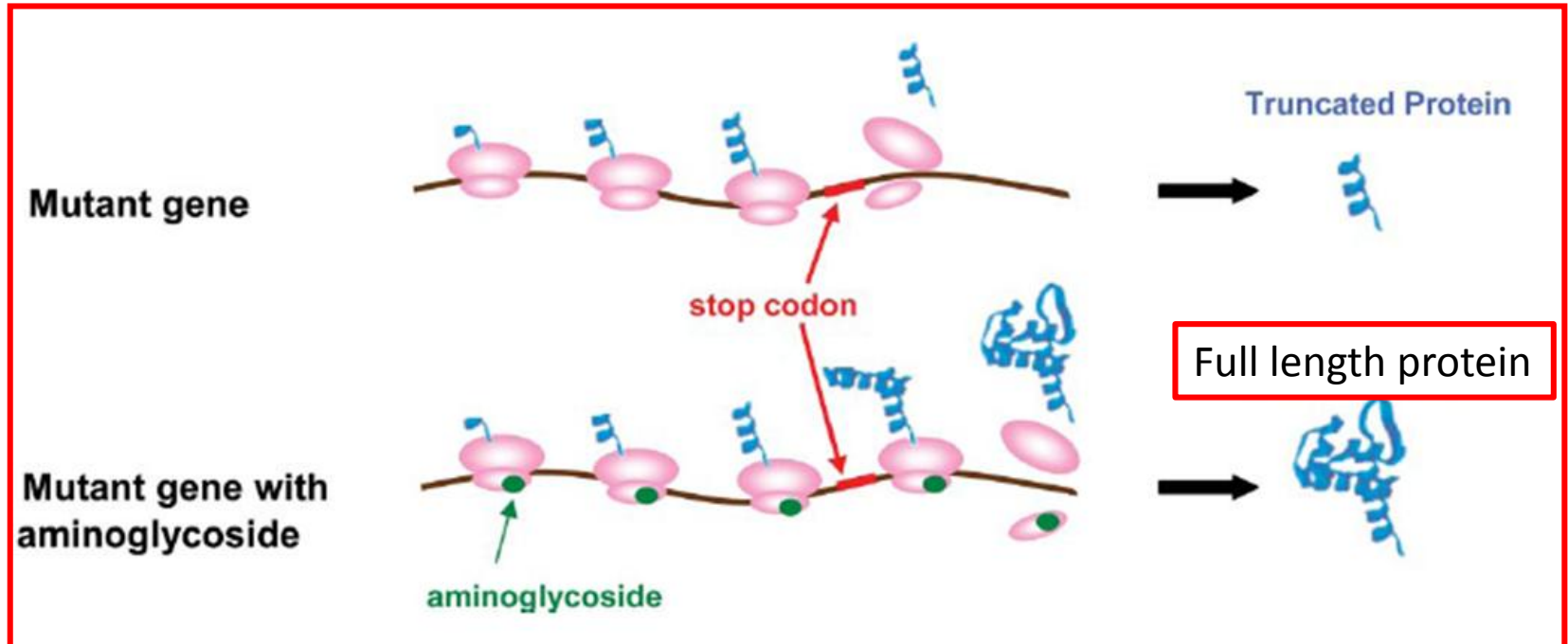
**Shortening ( ~20sec) of coagulation time**

## Molecular mechanisms for new therapeutic approaches (2)

### RNA directed/based therapy

- Engineered small RNA: RNAi, U1snRNA
- **Drug-induced translational readthrough of stop codons**
- Transcription activation TALE-TF

# mRNA-based approaches: Drug-induced translational readthrough of stop codons



Aminoglycosides bind the decoding site within the A-site in the ribosomal small subunit

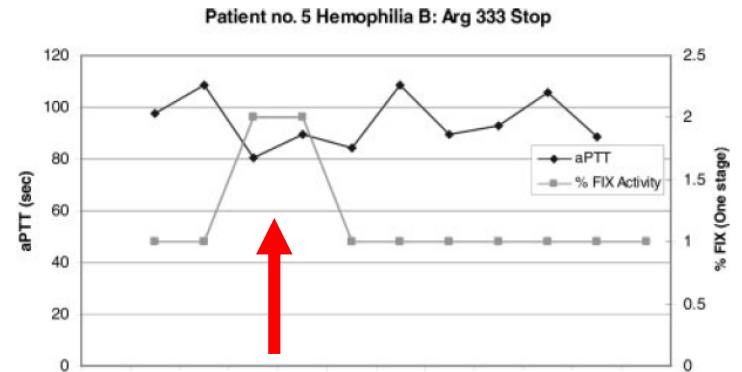
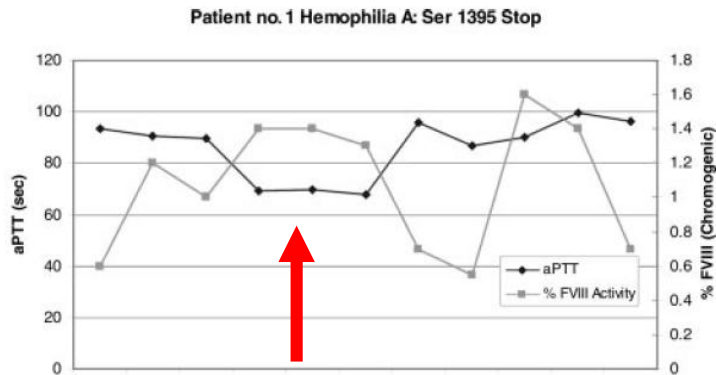
# Aminoglycoside suppression of nonsense mutations in severe hemophilia

Paula D. James, Sanj Raut, Georges E. Rivard, Man-Chiu Poon, Margaret Warner, Susan McKenna, Jayne Leggo, and David Lillicrap

BLOOD, 1 NOVEMBER 2005 · VOLUME 106, NUMBER 9

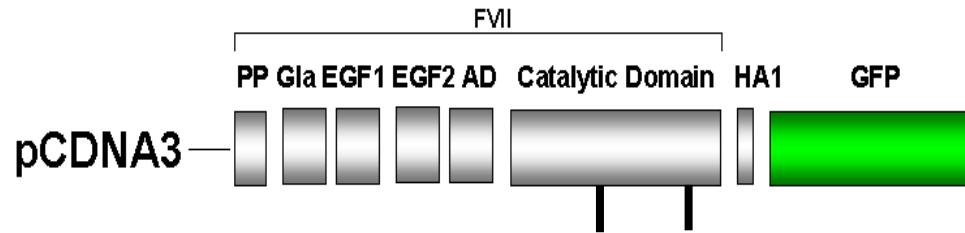
**Table 1. Patients with severe hemophilia with known nonsense mutations treated with gentamicin**

| Patient no. | Hemophilia | Nucleotide | Mutation    | Conserved amino acid |
|-------------|------------|------------|-------------|----------------------|
| 1           | A          | 4241C > A  | Ser1395Stop | No                   |
| 2           | B          | 30875C > T | Arg252Stop  | Yes                  |
| 3           | A          | 6403C > T  | Arg2116Stop | Yes                  |
| 4           | A          | 1536C > T  | Arg427Stop  | Yes                  |
| 5           | B          | 31118C > T | Arg333Stop  | Yes                  |

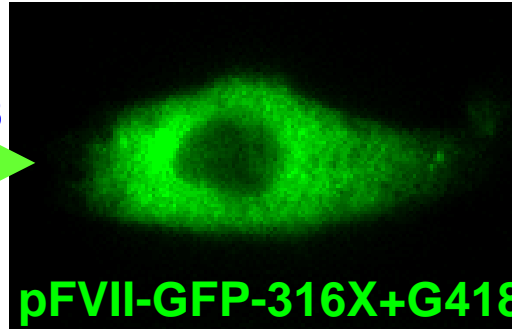




# FVII Deficiency: cellular models



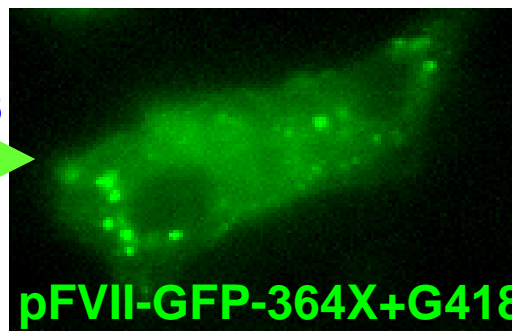
+ G418



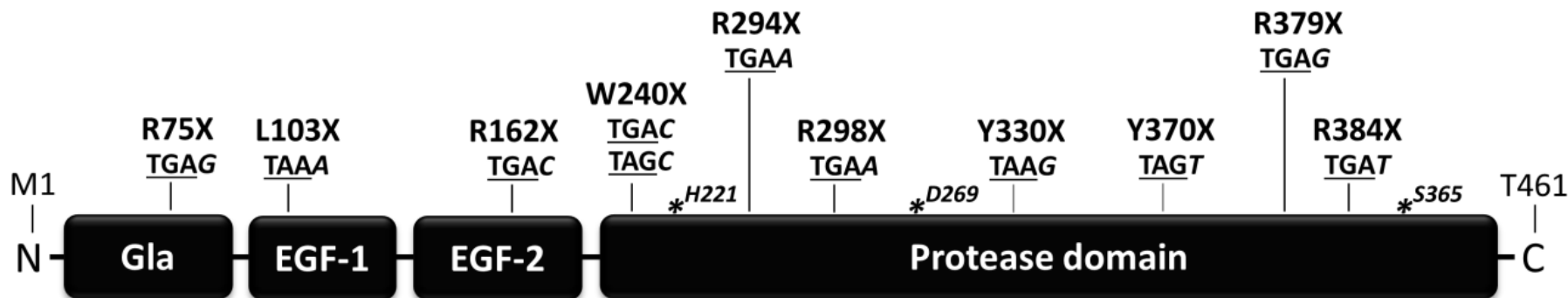
fluorescence supports partial rescue of full-length FVII biosynthesis



+ G418

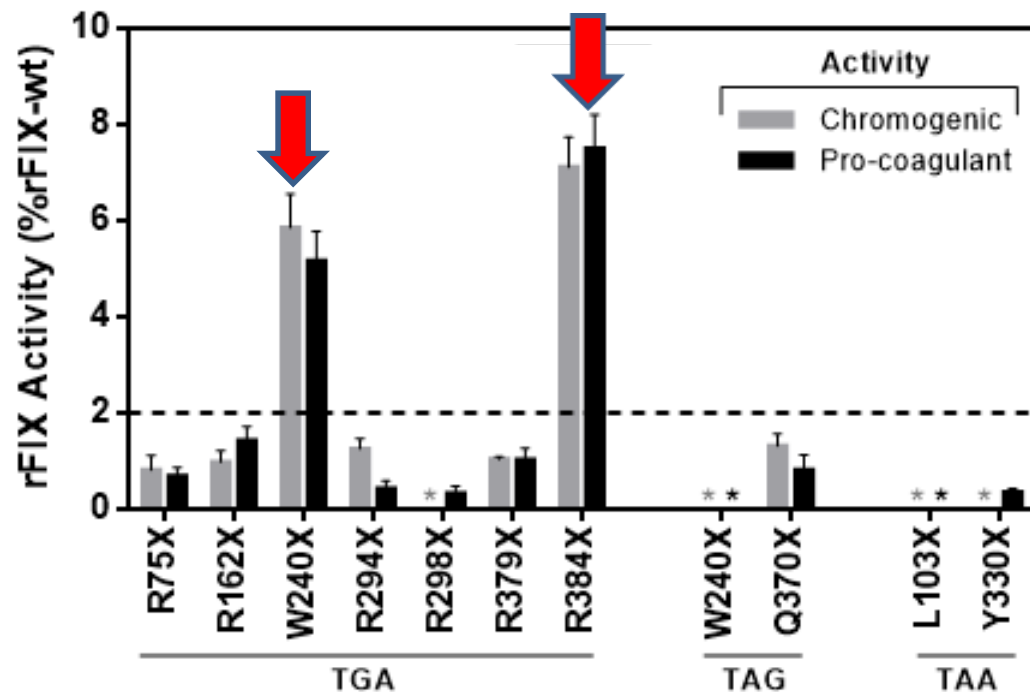


**Drug-induced translational readthrough investigated by expression of nonsense variants present in 70 % of HB patients with premature stop codons**



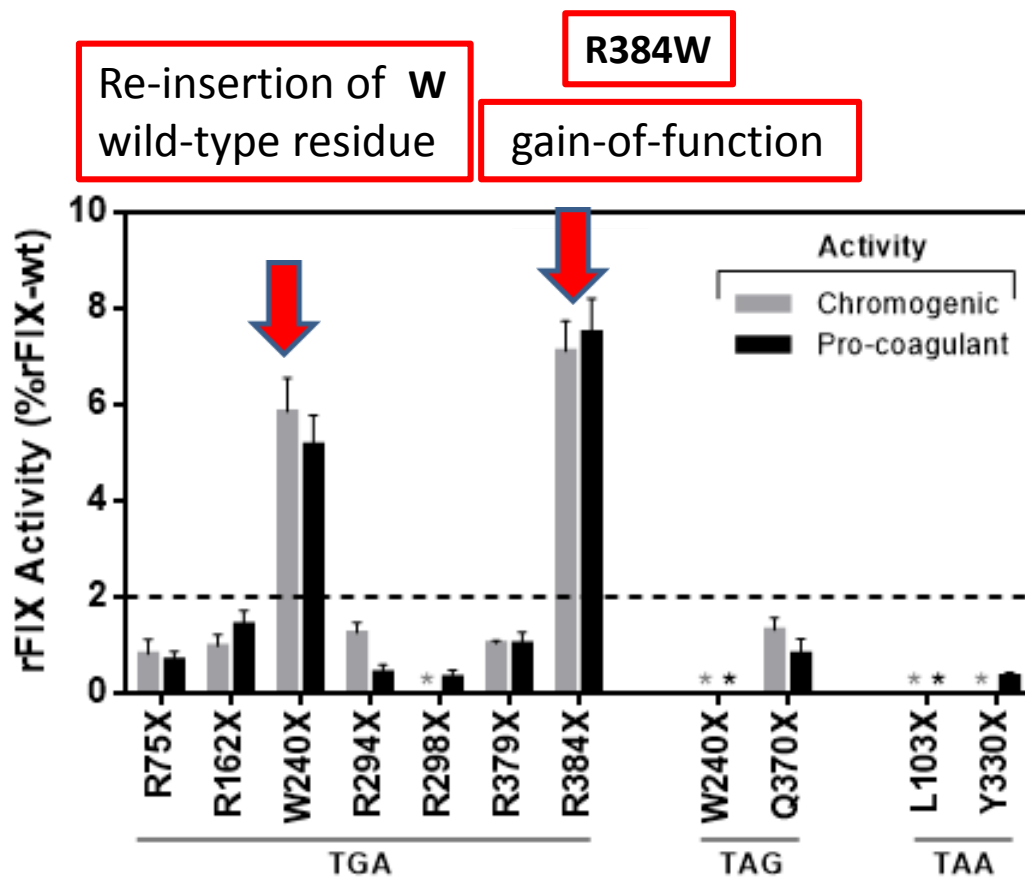

**Mattia Ferrarese**

only **two** variants displayed a **remarkable rescue of activity**

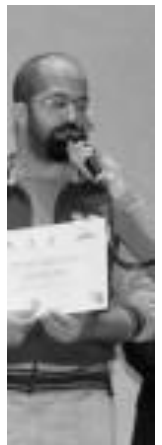
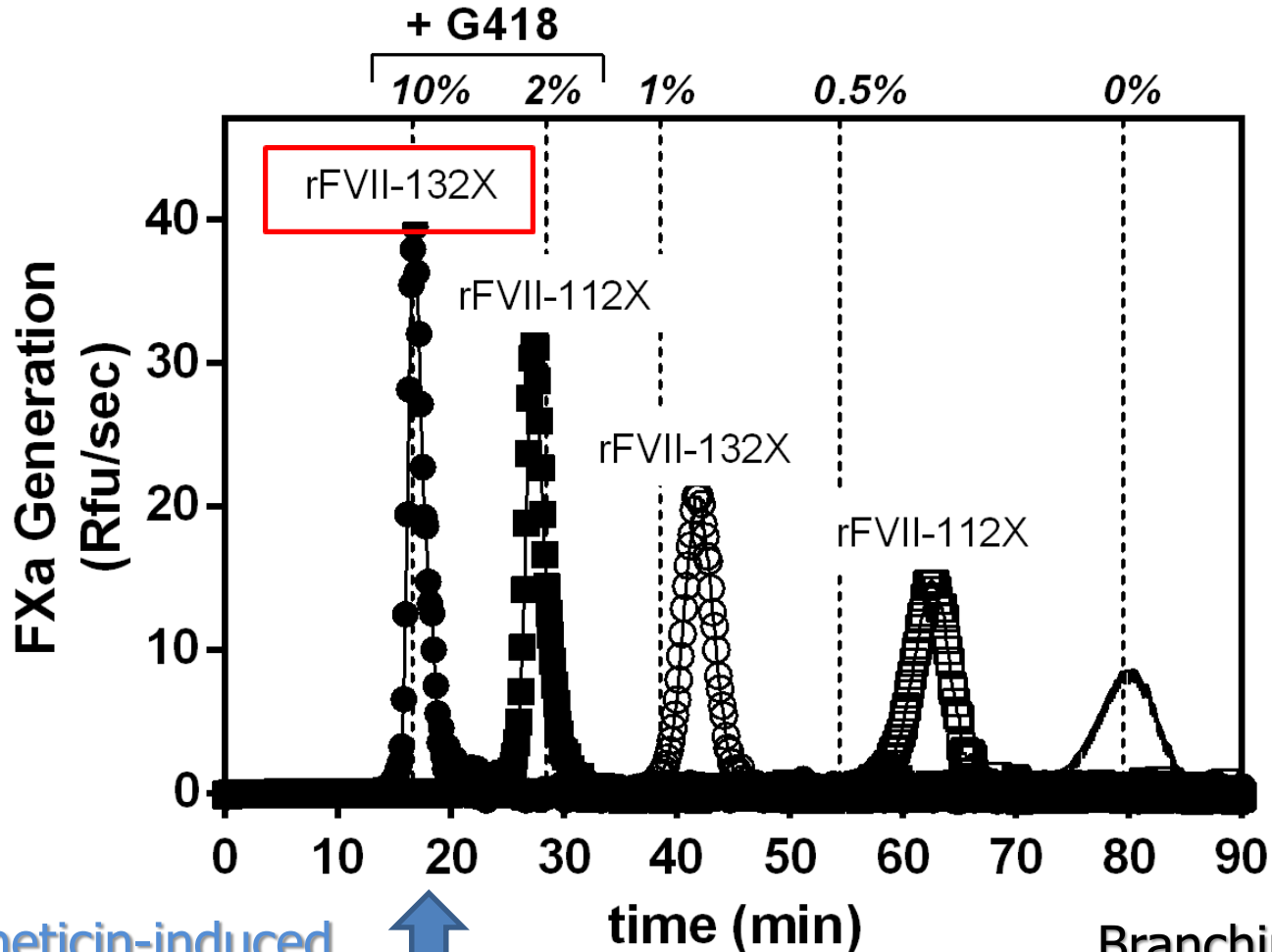


Expression of the **most probable missense variants** arising from readthrough

only **two** variants displayed a **remarkable rescue of activity**



Stop codons in FVII deficiency:  
the appreciable rescue of the p.Cys132X is driven  
by re-insertion of the wild-type residue!



Branchini  
et al JTH 2016

## Drug-induced translational readthrough of stop codons

- Only a few nonsense mutations in coagulation factor genes are remarkably responsive to drug-induced readthrough due to specific nucleotide sequence and protein structure constraints.
- The recombinant expression of nonsense variants helps interpreting the poor response reported in the few investigated patients  
Helps selecting candidate patients eligible for treatment

## Molecular mechanisms for new therapeutic approaches (2)

### RNA directed/based therapy

- Engineered small RNA: RNAi, U1snRNA
- Drug-induced translational readthrough of stop codons
- **Transcription activation - TALE-TF**



# AN ENGINEERED TALE-TRANSCRIPTION FACTOR RESCUES F7 PROMOTER ACTIVITY IMPAIRED BY MUTATIONS CAUSING SEVERE FACTOR VII DEFICIENCY



RNA



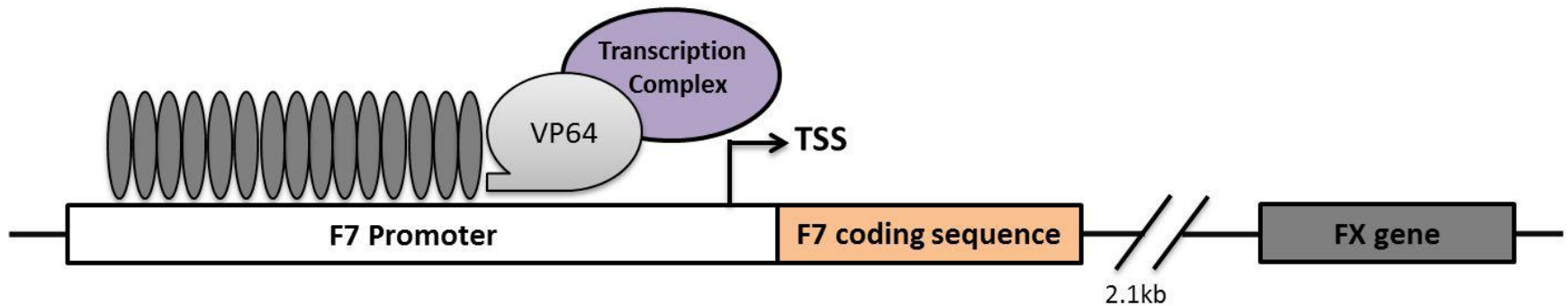
**Silvia Pignani**  
Department of Life Sciences and Biotechnology  
University of Ferrara-Italy

pgnslv@unife.it

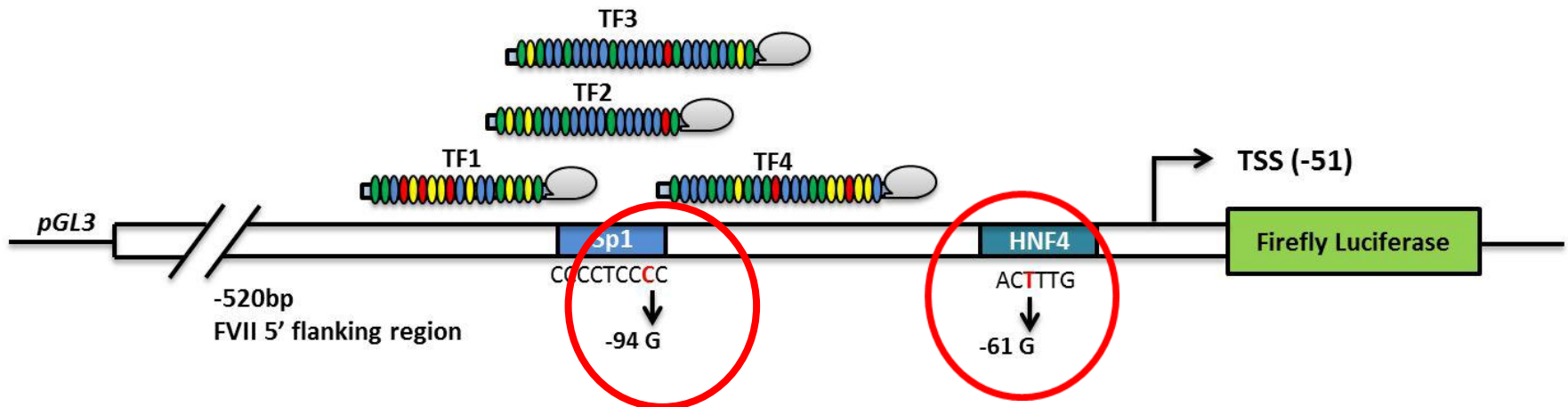


# ENGINEERED TALE-TRANSCRIPTION FACTORS TO RESCUE TRANSCRIPTION OF FACTOR VII IMPAIRED BY PROMOTER MUTATIONS

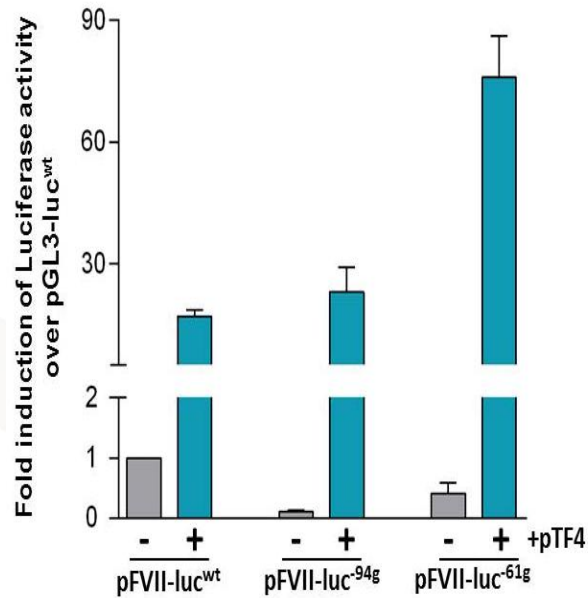
a



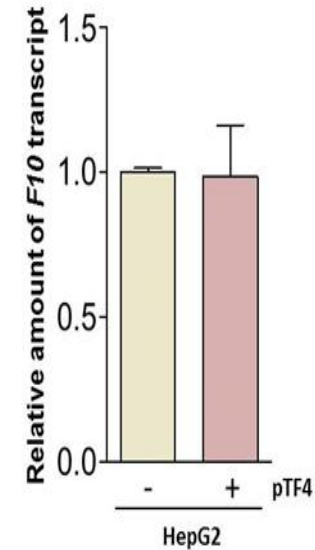
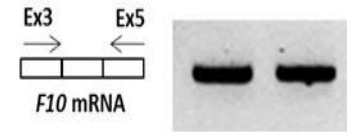
b



# TALE-TF4 enhances activity of the *F7* promoter in HepG2 cells

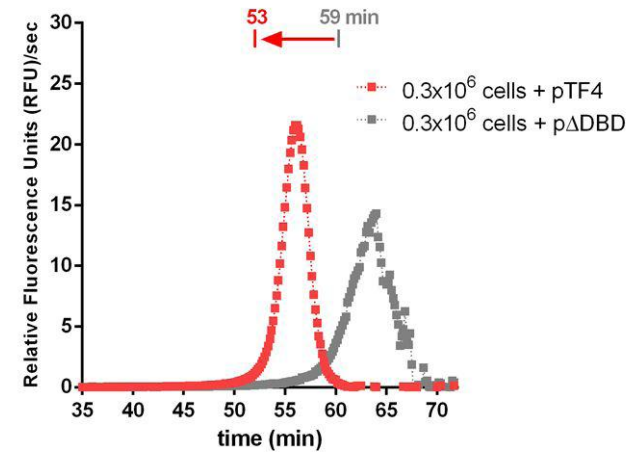
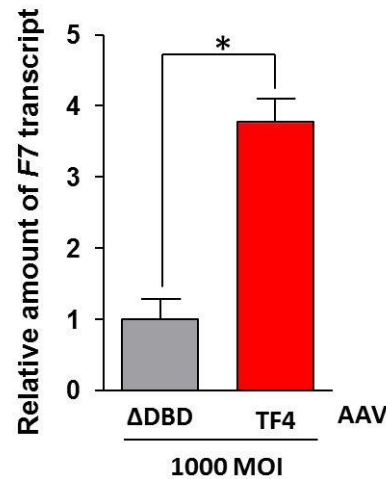
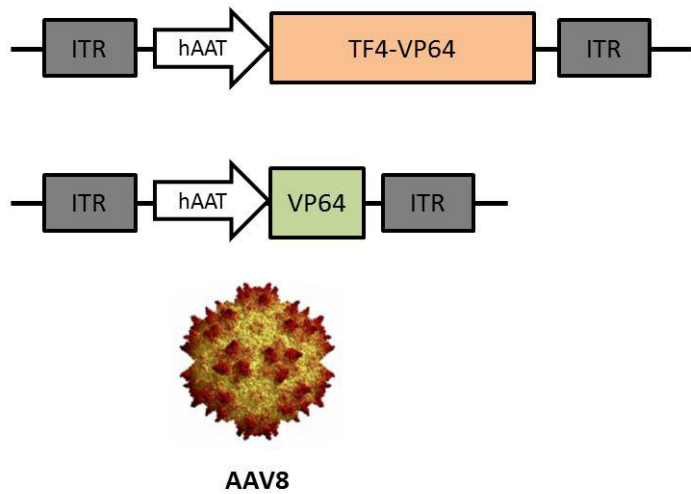


wild type mutants



TALE-TF4 does not induce  
transcription of *F10*

# TF4 increased F7 mRNA and FVII procoagulant levels in hepatocytes transduced by AAV



Experimental evidence for TALE-TFs as gene-specific tools useful to counteract disease-causing promoter mutations

## Gene Editing (Correction/Insertion):

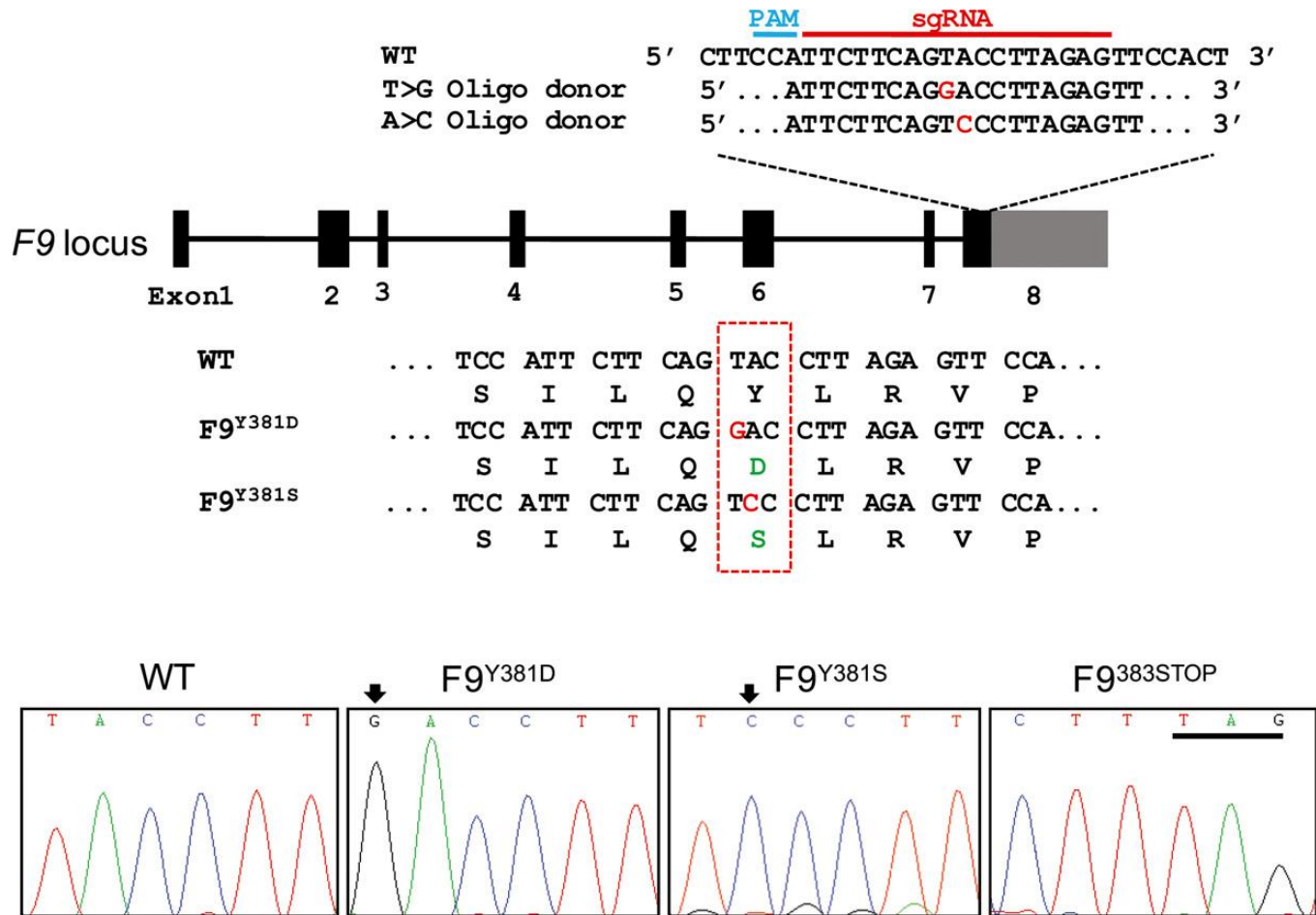
Zinc Finger Nuclease

TALE Nuclease

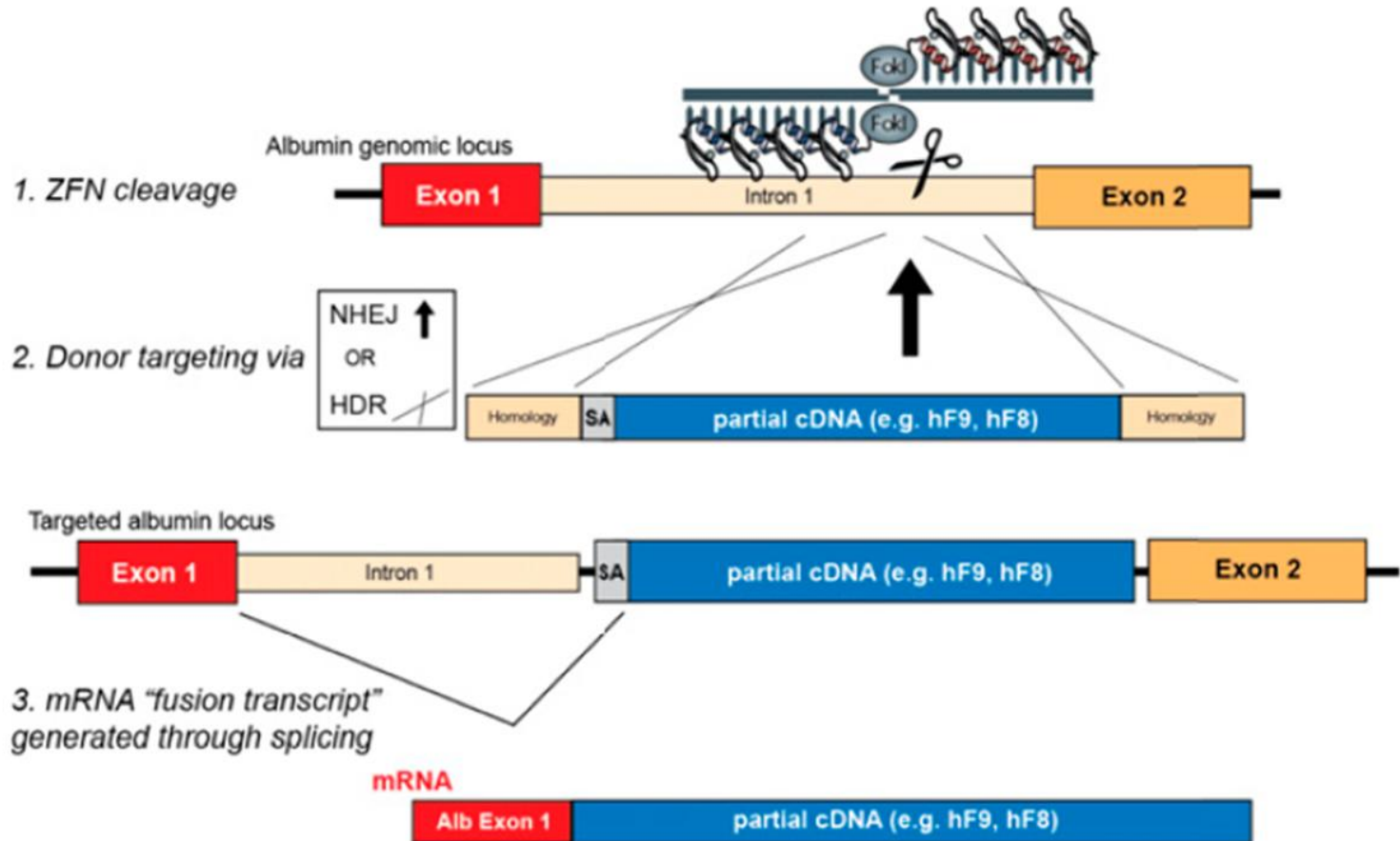
CRISPR/Cas9

Without nuclease

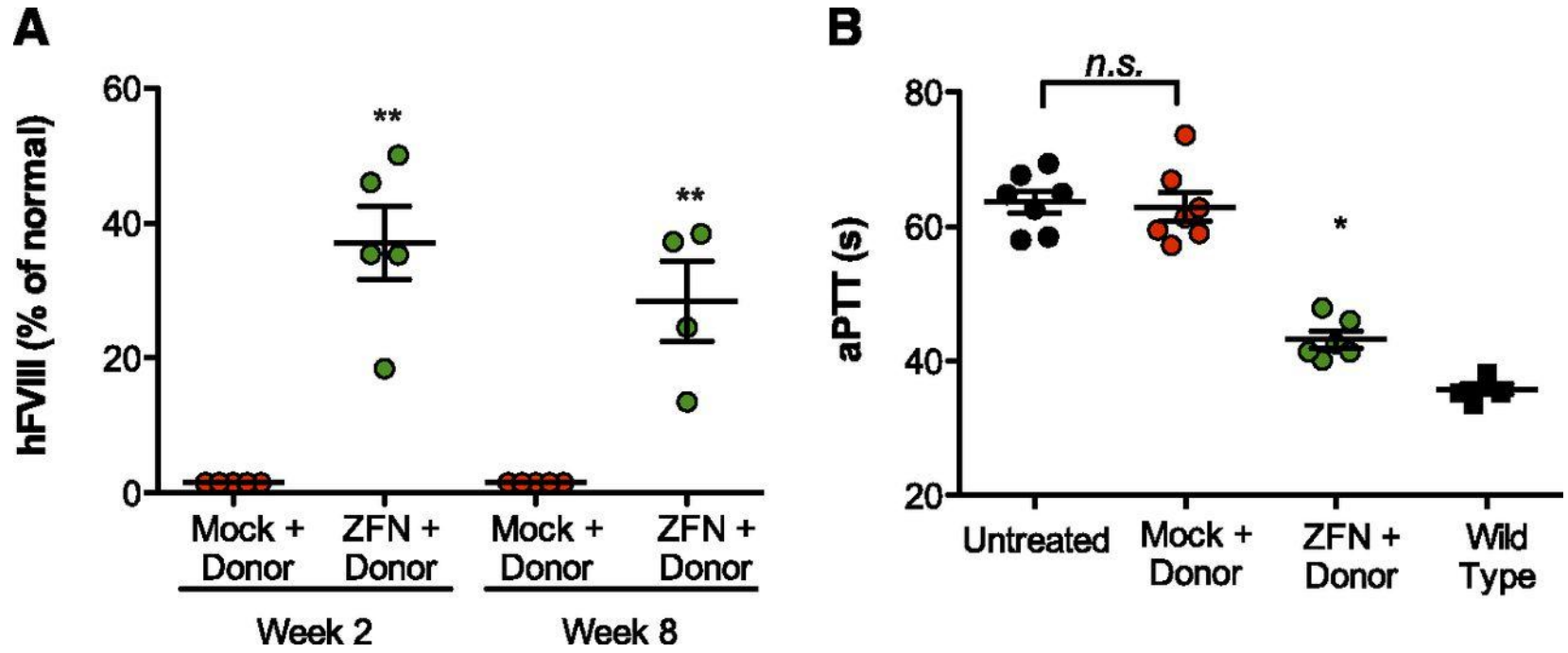
# Genome editing in situ by CRISPR/Cas9 restores hemostasis in F9 mutant mice



**Targeting albumin supports production of therapeutic levels of FVIII and functional correction of hemophilia A phenotype.**

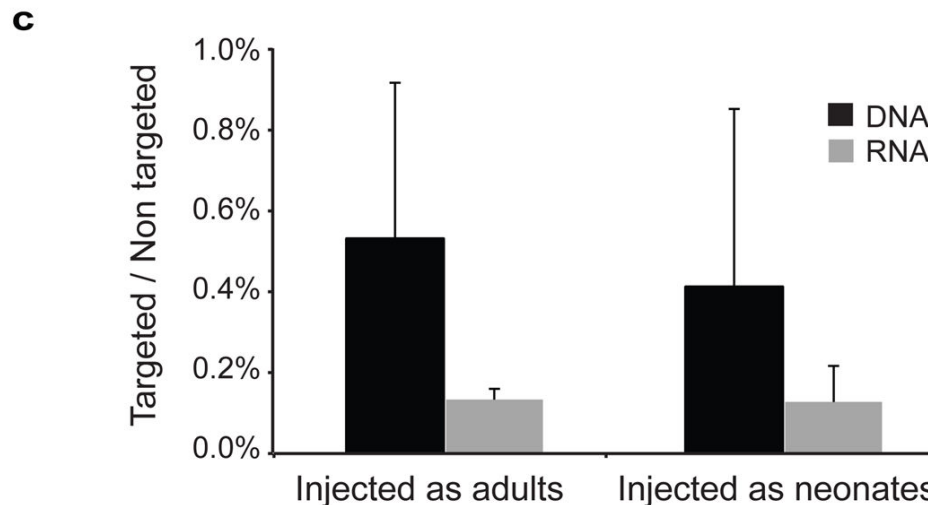
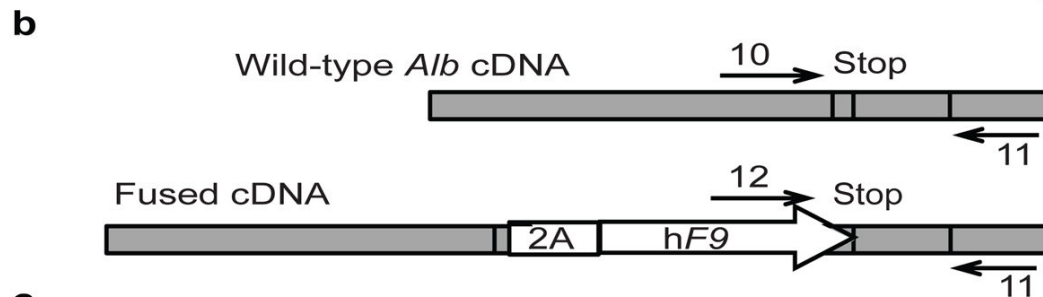
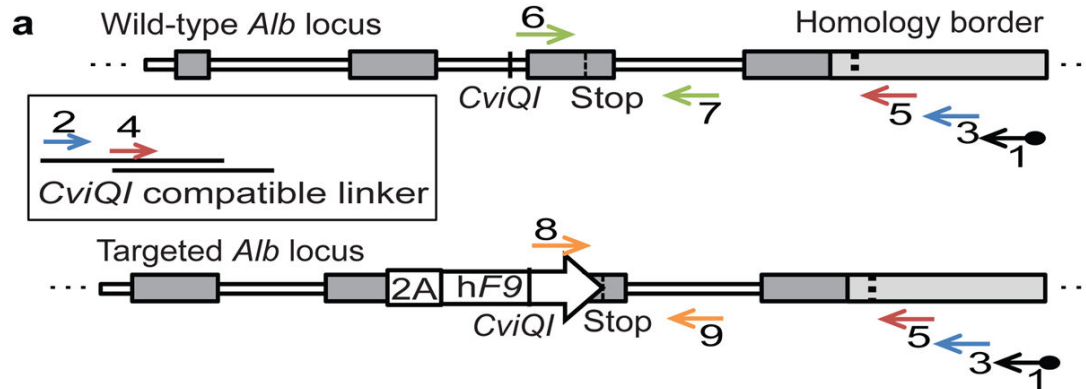


Targeting albumin supports production of therapeutic levels of FVIII and functional correction of hemophilia A phenotype.



Rajiv Sharma et al. Blood 2015;126:1777-1784

# Promoterless gene targeting without nucleases ameliorates haemophilia B in mice







**UNIVERSITÀ  
DEGLI STUDI  
DI FERRARA**  
- EX LABORE FRUCTUS -

**Mirko Pinotti**

